

RESEARCH

Open Access



Effect of a weight loss program on serum adiponectin and insulin resistance among overweight and obese premenopausal females

Walaa H. Foula^{1*} , Rana H. Emara¹, Mona K. Eldeeb², Samiha A. Mokhtar³ and Fikrat A. El-Sahn¹

Abstract

Background: Obesity has emerged as a public health crisis in many populations including Egypt. Adipose tissue produces a number of adipokines, one of them is adiponectin which has attracted much attention because of its antidiabetic and antiatherogenic effects.

Objective: To determine the effect of a weight loss program on serum adiponectin level and insulin resistance among overweight and obese adult premenopausal females.

Study design: A pre-postintervention study was carried out among 95 premenopausal overweight and obese females (body mass index ≥ 25 kg/m²) aged 20 to 40 years at the integrated health clinic affiliated to the High Institute of Public Health, Alexandria, Egypt, from February 2016 to February 2017. All participants underwent a weight loss program based on a reduced calorie balanced diet and advised to increase their physical activity. Dietary instructions and follow-up were done weekly throughout 16 weeks. Blood samples were collected to investigate serum adiponectin level and insulin resistance at the beginning and the end of the intervention.

Results: After 16 weeks, a significant decrease in body weight by 9.7% was associated with a significant increase in serum adiponectin from 13.3 ± 4.9 μ g/ml to 18.5 ± 5.6 μ g/ml. Both fasting insulin and insulin resistance had decreased significantly by 13.6% and 13.7%, respectively.

Conclusion: A weight reduction program depending on a reduced calorie diet for 16 weeks was associated with a significant increase in total adiponectin level and reduction in insulin resistance. An emphasis on the importance of keeping normal weight through nutritional education and the promotion of healthy diets is recommended to reduce the risk of occurrence of insulin resistance, type 2 diabetes, and cardiovascular diseases.

Keywords: Adipokines, Adiponectin, Insulin resistance, Weight reduction

* Correspondence: wfoula@gmail.com

¹Nutrition Department, High Institute of Public Health, Alexandria University, Alexandria, Egypt

Full list of author information is available at the end of the article



© The Author(s). 2020 **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

1 Introduction

Obesity is considered now a major public health problem. In 2014, more than 1.9 billion adults all over the world (about 39%) were overweight. Of these, over 600 million (about 13%) were obese [1]. According to the results of the Egypt Demographic and Health Survey 2014, 36.5% of women aged 15–49 years were overweight and 48.1% were obese [2]. The risk of diabetes, hypertension, and dyslipidemia increases with obesity.

Adipose tissue is a highly active endocrine organ that produces a number of hormones and other substances called “adipokines” [3], such as tumor necrosis factor (TNF- α), interleukin-6 (IL-6), leptin, and adiponectin [4]. Adiponectin has a special role because of its antidiabetic and antiatherogenic effects [5]. Plasma adiponectin levels in humans range from 3 to 30 $\mu\text{g/ml}$ [5]. It accounts for 0.01% of the total human plasma proteins, and this makes it the most abundant adipose tissue protein [6]. Plasma levels were significantly lower in men than women [7], obese subjects [8], metabolic syndrome [9] and type 2 diabetic patients [10], and those with coronary artery disease [10].

Caloric reduction is an easy and cost-effective measure in the management of obesity and comorbidities [11, 12]. Studies comparing adiponectin levels after weight loss programs are inconclusive [13–15]. The aim of the present study was to investigate the effect of a 16-week weight reduction program depending on a balanced low-calorie diet on serum total adiponectin level and insulin resistance among a group of overweight and obese premenopausal females.

2 Methods

2.1 Study design

One group pre-post intervention study design was carried out. The field study was conducted from February 2016 to February 2017.

2.2 Study setting

The study was carried out in the integrated health clinic affiliated to the High Institute of Public Health (HIPH), Alexandria, Egypt.

2.3 Participants and sample size

All subjects were premenopausal, nonpregnant, nonlactating overweight and obese ($\text{BMI} \geq 25 \text{ kg/m}^2$) adult females aged 20 to 40 years. They showed no evidence of heart, renal, liver, cancer, diabetes mellitus, or any other medical disorders, or history of surgeries for obesity, or taking medications (oral steroids, thiazolidinediones, angiotensin-converting enzyme inhibitors, angiotensin II receptor blockers, clonidine like sympathoinhibitory antihypertensive agent and fenofibrate).

The study started by 122 overweight and obese adult females and ended by 95, and the dropout rate was 22%. A total sample of 95 overweight and obese adult females was required to estimate the effect size of the weight loss program for change at adiponectin level = 0.30, using alpha error = 0.05 and dropout rate = 10% and baseline adiponectin level = $2.98 \pm 1.0 \mu\text{g/ml}$ [16]. This sample size was calculated using G.power software. Two days of the week were selected, and all subjects who attended the study setting and were fulfilling the selection criteria at these two days were included in the study until the required sample size was reached.

2.4 Data collection

2.4.1 Pre-intervention assessment phase

1. A predesigned interviewing questionnaire was used at the beginning of the study to collect personal data, medical history, weight history, physical activity, and dietary habits.
2. Twenty-four-hour dietary recall technique was carried out as a dietary assessment tool.
3. Anthropometric measurements, height (Ht), weight (Wt), waist circumference (WC), and hip circumference (HC), were measured [17]. Body mass index (BMI) was calculated as weight (kg)/height (m)². Waist-hip ratio (WHR) was calculated by dividing the waist circumference by the hip circumference. Waist-height ratio (WHtR) was calculated by dividing the waist circumference by the measured height in centimeters.
4. Assessment of total fat mass, skeletal muscle mass, and total body water was done using InBody720 which depends on the bioelectrical impedance analysis (BIA) technique [18], then body fat percentage, skeletal muscle percentage, and body water percentage were calculated.
5. Laboratory investigations were carried out where blood samples were collected in heparinized syringes after 8 h overnight fast. Fasting serum glucose [19] and fasting serum insulin levels [20] were determined. Quantitative determination of the insulin level was performed using an enzyme-linked immunosorbent assay (ELISA) kit supplied from DRG instruments GmbH, Germany (EIA-2935). The degree of insulin resistance was calculated by the updated computer homeostasis model assessment (HOMA2) [21] and referred to as homeostasis model assessment for insulin resistance (HOMA-IR). Serum samples were stored at -20°C unit until serum total adiponectin concentration was measured using commercial ELISA (human adiponectin ELISA kit; Boster Immunoleader, Pleasanton, CA, catalog number EK0595) [22].

2.4.2 Intervention phase

Weight loss program: All subjects were instructed to eat a well-balanced diet aiming at reducing 500–800 kcal/day less than individually calculated energy requirements, with the goal to achieve a rate of weight loss of 0.5–1.0 kg/week [23]. Resting energy expenditure (REE) was calculated by the Mifflin St Jeor equation [24], then it was multiplied by a coefficient of correction for physical activity level to get energy requirements. The diet was designed to provide 20–35% of energy derived from fat, 45–65% from carbohydrates, and 10–35% from proteins [25]. Subjects were advised to exercise or walk 30 min per day, 5 days a week.

2.4.3 Follow-up phase

Dietary instructions were reinforced and monitored weekly throughout the 16-week intervention period. Compliance to physical activity was recorded weekly. The females were classified according to their compliance to physical exercise recommendation into three categories: no physical exercise, irregular physical exercise, and physical exercise as recommended.

2.4.4 Evaluation phase

At the end of the 16-week intervention period, the outcomes of the intervention were evaluated through re-assessment of changes in the anthropometric measurements, body components, and laboratory investigations.

2.5 Ethical consideration

Approval of the Ethics Committee of HIPH, Alexandria University, Egypt, was obtained on 9 February 2016. All patients were informed, and a written consent was

obtained from all participants after explaining the aim of the study.

2.6 Data management and statistical analysis

Data were managed and analyzed using the statistical software IBM SPSS version 20. All statistical analyses were done using a two-tailed test and alpha error 0.05. Numeric data was tested for normality using the Kolmogorov-Smirnov test. Normally distributed data were presented as mean \pm standard deviation (SD). Student's *t* test, one-way analysis of variance (ANOVA), paired *t* test, simple correlation coefficient, and multiple linear regression were used in analyzing data [26].

3 Results

The mean age of the sample was 31.12 ± 7.18 years. The anthropometric and laboratory characteristics of the studied sample before and after the intervention are presented in Table 1. After the intervention, body weight and BMI decreased significantly by 9.7% ($P < 0.0001$) and also WC decreased significantly by 8.2% ($P < 0.0001$). WHR and WHtR decreased significantly by 2.7% and 8.2%, respectively ($P < 0.0001$). The changes in the body composition with the interventions indicated a significant decrease in the percent of body fat by 7.8% ($P < 0.0001$) and a significant increase in the percent of body muscle and body water by 6% and 6.9%, respectively ($P < 0.0001$). The laboratory changes after the intervention demonstrated a significant increase in adiponectin by 50.2% ($P < 0.0001$) from 13.3 ± 4.9 $\mu\text{g/ml}$ to 18.5 ± 5.6 $\mu\text{g/ml}$. Both fasting insulin and HOMA-IR had decreased significantly by 13.6% and 13.7%, respectively ($P < 0.0001$), but there was a slight insignificant decrease in fasting blood glucose (0.2%).

Table 1 Anthropometric and laboratory characteristics of the sample of overweight and obese women before and after the intervention ($n = 95$), Alexandria, Egypt, 2016-2017

| Characteristics | Before, mean \pm SD | After, mean \pm SD | Percent of change (%) | t_d | <i>P</i> value |
|--|-----------------------|----------------------|-----------------------|--------|----------------|
| Weight, kg | 94.4 \pm 18.5 | 85.1 \pm 16.9 | -9.7 | 18.185 | < 0.001* |
| Body mass index, kg/m ² | 35.8 \pm 6.9 | 32.3 \pm 6.2 | -9.7 | 18.118 | < 0.001* |
| Waist circumference, cm | 100.9 \pm 12.4 | 92.6 \pm 11.5 | -8.2 | 17.425 | < 0.001* |
| Waist-hip ratio (WHR) | 1.03 \pm 0.07 | 1.00 \pm 0.08 | -2.7 | 5.056 | < 0.001* |
| Waist-height ratio (WHtR) | 0.62 \pm 0.079 | 0.57 \pm 0.075 | -8.2 | 17.609 | < 0.001* |
| Body fat percent, % | 47.4 \pm 4.4 | 43.7 \pm 5.4 | -7.8 | 12.837 | < 0.001* |
| Body muscle percent, % | 29.0 \pm 2.3 | 30.7 \pm 2.8 | +6.0 | 9.742 | < 0.001* |
| Body water percent, % | 38.6 \pm 3.1 | 41.2 \pm 3.9 | +6.9 | 13.124 | < 0.001* |
| Serum total adiponectin ($\mu\text{g/ml}$) | 13.3 \pm 4.9 | 18.5 \pm 5.6 | +50.2 | 8.701 | < 0.001* |
| Fasting serum glucose (mg/dl) | 89.9 \pm 12.1 | 88.8 \pm 12.1 | -0.22 | 0.813 | 0.418 |
| Fasting serum insulin ($\mu\text{IU/ml}$) | 10.3 \pm 5.6 | 7.7 \pm 3.3 | -13.6 | 6.376 | < 0.001* |
| HOMA-IR | 1.3 \pm 0.7 | 1.0 \pm 0.4 | -13.7 | 6.338 | < 0.001* |

SI conversion factors: to convert glucose from mg/dl to mmol/l, multiply by 0.0555; and insulin from $\mu\text{IU/ml}$ to pmol/l, multiply by 7.175

Abbreviations: t_d *t* Difference in paired *t* test, HOMA-IR Homeostasis model assessment for insulin resistance

*Statistically significant

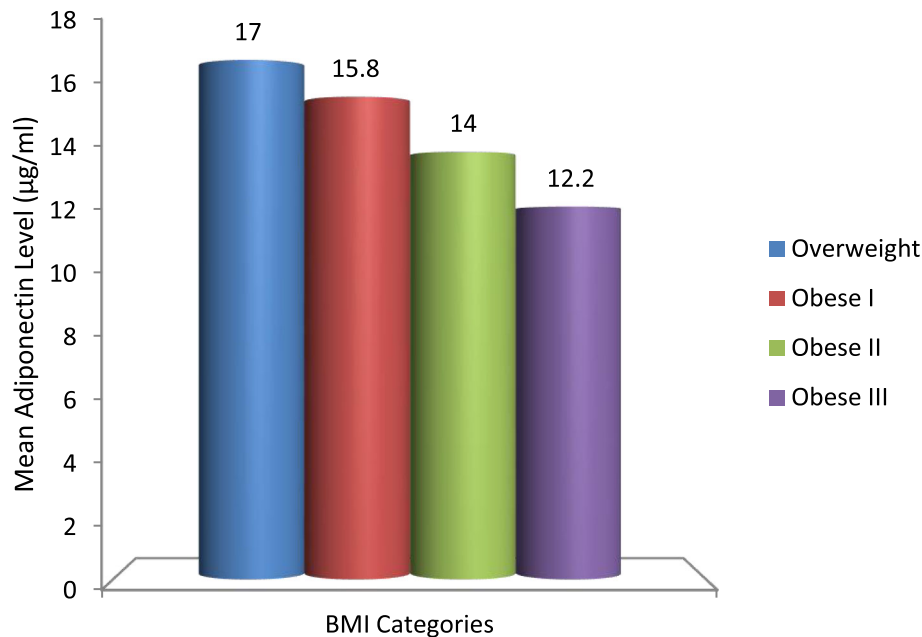


Fig. 1 The mean adiponectin level among the studied sample according to BMI categories

An inverse relationship between the mean of the adiponectin level and BMI was observed. The mean adiponectin level among females with overweight was higher than the level of those with grade III obesity (17.0 µg/ml and 12.2 µg/ml, respectively), and also the mean adiponectin level among females with grade I obesity (15.8 µg/ml) was higher than the level of those with grade III obesity, and both differences were statistically significant ($F = 3.817$, $P = 0.013$) (Fig. 1).

The correlation coefficients of total adiponectin, HOMA-IR before the intervention, and some

anthropometric and laboratory characteristics are shown in Table 2. There was a significant direct correlation between adiponectin level and both body muscle percentage ($r = 0.302$) and body water percentage ($r = 0.317$), while there was a significant indirect correlation between adiponectin level and each of WHR ($r = -0.393$), body fat percentage ($r = -0.308$), body weight ($r = -0.204$), fasting insulin ($r = -0.210$), and HOMA-IR ($r = -0.195$). On the other hand, there was a significant direct correlation between HOMA-IR and each of body weight ($r = 0.474$), BMI ($r = 0.435$), WC ($r = 0.330$), WHtR ($r =$

Table 2 Correlations of anthropometric and laboratory characteristics with serum total adiponectin and HOMA-IR before the intervention ($n = 95$)

| Characteristics | Serum total adiponectin | P value | HOMA-IR | P value |
|---------------------------|-------------------------|---------|---------------------|---------|
| Body weight | -0.204 ^a | 0.047 | 0.474 ^a | < 0.01 |
| Body mass index | -0.119 | 0.249 | 0.435 ^a | < 0.01 |
| Waist circumference | -0.103 | 0.318 | 0.330 ^a | < 0.01 |
| Waist-hip ratio (WHR) | -0.393 ^a | 0.001 | 0.186 | 0.071 |
| Waist-height ratio (WHtR) | -0.033 | 0.751 | 0.278 ^a | 0.006 |
| Body fat percentage | -0.308 ^a | 0.002 | 0.226 ^a | 0.028 |
| Body muscle percentage | 0.302 ^a | 0.003 | -0.147 | 0.155 |
| Body water percentage | 0.317 ^a | 0.002 | -0.226 ^a | 0.028 |
| Fasting serum glucose | 0.170 | 0.099 | 0.393 ^a | < 0.01 |
| Fasting serum insulin | -0.210 ^a | 0.041 | 0.998 ^a | < 0.01 |
| HOMA-IR | -0.195 ^a | 0.050 | N/A | - |
| Serum total adiponectin | N/A | - | -0.195 ^a | 0.050 |

Abbreviations: HOMA-IR Homeostasis model assessment for insulin resistance, N/A Not applicable

^aStatistically significant

Table 3 Multiple regression analysis of variables associated with adiponectin level before the intervention

| Factor | B | SE | Beta | t | Sig. |
|---|----------|-------|---------|---------|---------|
| History of practicing physical activity | 0.552 | 0.165 | 0.294 | 3.341 | < 0.01* |
| Number of main meals | 0.934 | 0.418 | 0.207 | 2.232 | 0.028* |
| Body mass index | - 0.259 | 0.097 | 0.366 | - 2.668 | 0.009* |
| Waist-hip ratio | - 22.836 | 6.687 | - 0.339 | - 3.415 | < 0.01* |
| Percent body fat | - 0.343 | 0.152 | - 0.307 | - 2.254 | 0.027* |
| HOMA-IR | - 1.545 | 0.714 | - 0.229 | - 2.163 | 0.033* |

$R^2 = 36.1\%$

$F = 7.017, P = 0.0001^*$

*Statistically significant

0.278), and body fat percentage ($r = 0.226$), while there was a significant indirect correlation between HOMA-IR and body water percentage ($r = - 0.226$).

Two multiple regression analysis models (multivariable) to uncover factors which might be related to adiponectin level and HOMA-IR among the studied sample before intervention were done (Tables 3 and 4). WHR ($B = - 22.836, SE B = 6.687, P = 0.001$), body fat percentage ($B = - 0.343, SE B = 0.152, P = 0.027$), and HOMA-IR ($B = - 1.545, SE B = 0.714, P = 0.033$) were independently, negatively, and significantly associated with serum total adiponectin level [the unstandardized beta (B), the standard error for the unstandardized beta ($SE B$), and the probability value (P)] (Table 3), while only BMI ($B = 0.029, SE B = 0.010, P = 0.003$) was independently, positively, and significantly associated with HOMA-IR (Table 4).

The correlation coefficients between differences in each of adiponectin level, HOMA-IR, and some anthropometric and laboratory characteristics of the sample are shown in Table 5. There was no statistically significant correlation between the increment in adiponectin level and the change in any anthropometric or laboratory parameters. A significant direct correlation between the difference in HOMA-IR and difference in each of WHR ($r = 0.310$), body weight ($r = 0.238$), BMI ($r = 0.229$), and WC ($r = 0.202$) was found.

A multiple regression analysis to uncover factors which might be related to the change in HOMA-IR was done. The model explained 22.3% of the variation in the difference of HOMA-IR, and it was statistically

Table 4 Multiple regression analysis of variables associated with HOMA-IR before the intervention

| Factor | B | SE | Beta | t | Sig. |
|---------------------------|-------|-------|-------|-------|--------|
| Family history of obesity | 0.399 | 0.161 | 0.245 | 2.478 | 0.015* |
| Number of main meals | 0.187 | 0.070 | 0.257 | 2.669 | 0.009* |
| Body mass index | 0.029 | 0.010 | 0.280 | 3.036 | 0.003* |

$R^2 = 44.0\%$

$F = 12.582, P = 0.0001^*$

*Statistically significant

significant ($F = 5.103, P = 0.0001$). Body mass index ($B = 0.095, SE B = 0.029, P = 0.002$) and WHR ($B = 2.746, SE B = 0.989, P = 0.007$) were independently, positively, and significantly associated with the change in HOMA-IR (Table 6).

Patients were classified according to compliance to physical activity recommendations into three categories as shown in Table 7. There was no statistically significant difference between the mean differences of any anthropometric measurements, adiponectin level, or insulin resistance among the three categories of the sample.

4 Discussion

In this study, the hypothesis that a short-term (16 weeks) weight reduction program depending mainly on low caloric balanced diet can change serum total adiponectin level and insulin resistance was tested. In contrast to other adipokines and, although it is produced by adipocytes, adiponectin level is paradoxically lower in obese subjects than in non-obese subjects as revealed in a previous study [8]. This finding suggests that adipose tissue may exert a negative feedback on adiponectin production. In the present study, the mean total plasma adiponectin level was slightly higher than expected hypoadiponectinemia associated with overweight and obesity. This finding may be due to the wide range of normal reference (3–30 $\mu\text{g/ml}$) [5].

Results concerning changes in adiponectin level after weight loss in many studies are inconsistent. Several studies demonstrated no change in total plasma adiponectin after a short-term weight reduction intervention despite a significant reduction in body weight [27–29]. On the other hand, large weight reduction following bariatric surgery was accompanied by an increase in plasma adiponectin level [30, 31]. It has been demonstrated from the present study that a moderate weight loss of 9.3 kg (represented 9.7% of the original weight) induced by a balanced low caloric diet was accompanied by a significant increase in serum total adiponectin level from 13.3 to 18.5 $\mu\text{g/ml}$, which represented 50.2% increase

Table 5 Correlations of anthropometric and laboratory characteristics with change (after-before) in serum total adiponectin and HOMA-IR ($n = 95$)

| | Change in serum total adiponectin | P value | Change in HOMA-IR | P value |
|-----------------------------------|-----------------------------------|---------|--------------------|---------|
| Change in body weight | -0.106 | 0.304 | 0.238 ^a | 0.020 |
| Change in body mass index | -0.095 | 0.362 | 0.229 ^a | 0.026 |
| Change in waist circumference | -0.099 | 0.342 | 0.202 ^a | 0.049 |
| Change in waist-hip ratio | -0.020 | 0.847 | 0.310 ^a | 0.002 |
| Change in waist-height ratio | -0.095 | 0.357 | 0.200 | 0.052 |
| Change in body fat percentage | -0.003 | 0.975 | 0.129 | 0.214 |
| Change in fasting serum glucose | 0.068 | 0.511 | 0.187 | 0.070 |
| Change in fasting serum insulin | -0.081 | 0.433 | 0.996 ^a | < 0.001 |
| Change in HOMA-IR | -0.093 | 0.372 | N/A | - |
| Change in serum total adiponectin | N/A | - | -0.093 | 0.372 |

Abbreviations: HOMA-IR Homeostasis model assessment for insulin resistance, N/A Not applicable

^aStatistically significant

from the original level and a decrease in insulin resistance by 13.7%.

The change in adiponectin level was not significantly correlated with the change in insulin sensitivity or the improvement in obesity parameters; this could be explained by the short period of the intervention. Meanwhile, the decline in insulin resistance was positively correlated to the decline in body weight, BMI, WC, and WHR. Regression analysis revealed that difference in BMI and difference in WHR were the two predictors for the improvement in insulin resistance index.

The relation between adiponectin level and BMI is inconsistent. Many studies reported a negative association between them [32–34], while another study demonstrated no association [35]. The present study revealed no significant correlation between them. Previous studies reported a negative correlation between adiponectin level and waist circumference as a parameter for increased abdominal fat accumulation [36, 37]. In the current study, we investigated three parameters (WC, WHR, and WHtR) to indicate the relation between adiponectin level and abdominal obesity. Of these parameters, WHR was the most appropriate indicator, as it was negatively related to adiponectin level, and was one of the predictors of adiponectin level in the multiple regression model.

Table 6 Multiple regression analysis of variables associated with change (after-before) in HOMA-IR

| Factor | B | SE | Beta | t | Sig. |
|-------------------------------|-------|-------|-------|-------|--------|
| Difference in BMI | 0.095 | 0.029 | 0.352 | 3.224 | 0.002* |
| Difference in waist-hip ratio | 2.746 | 0.989 | 0.297 | 2.777 | 0.007* |

$R^2 = 22.3\%$

$F = 5.103, P = 0.0001^*$

*Statistically significant

Apart from negative correlations with adiposity measures, studies revealed that adiponectin levels seem to be reduced prior to the development of type 2 diabetes, even after adjusting for measures of obesity, and that administration of adiponectin has been accompanied by increased insulin sensitivity [38, 39]. The present study extends findings of these studies by providing an example of an obesity-independent association of insulin resistance with adiponectin levels, as there was a significant indirect correlation between HOMA-IR and mean level of total adiponectin; also, a multiple regression analysis revealed HOMA-IR to be one of the significant independent predictors of total adiponectin level.

Regarding physical exercise as a part of the weight reduction program, a previous study reported that a weight loss program that included exercise increased the plasma adiponectin levels of obese women in a randomized trial [40]. In the present study, there was no statistically significant difference between the change in the mean adiponectin level or the mean of any anthropometric parameters, percent of body components, or insulin sensitivity among the three categories of the sample regarding physical activity compliance. This finding supports that the low caloric diet represented the more effective part of the intervention program; also, it raises the suggestion that more time is needed for the physical activity to affect the adiponectin level or cause a change in anthropometric measurements or insulin sensitivity.

4.1 Limitations of the study

There were certain limitations in undertaking this study; first, the design adopted in the study was one arm design lacking the control limb. Although this study design is considered a weak one, for ethical considerations, it was unethically to take a group of obese patients as a control group without doing any intervention for them despite

Table 7 Mean difference of anthropometric and laboratory characteristics before and after the intervention according to compliance with physical exercise

| Mean difference in the variables (after-before) intervention | No exercise, n = 41 | Irregular exercise, n = 44 | Regular exercise, n = 10 | Sig. F | P value |
|--|---------------------|----------------------------|--------------------------|--------|---------|
| Weight (kg) | -9.48 | -9.59 | -7.23 | 0.963 | 0.385 |
| Body mass index, kg/m ² | -3.58 | -3.65 | -2.75 | 0.947 | 0.392 |
| Waist circumference, cm | -8.32 | -8.66 | -6.90 | 0.576 | 0.564 |
| Waist-hip ratio (WHR) | -0.032 | -0.026 | -0.029 | 0.096 | 0.908 |
| Waist-height ratio (WHtR) | -0.051 | -0.053 | -0.043 | 0.565 | 0.570 |
| Body fat percentage | -3.27 | -4.20 | -2.77 | 1.786 | 0.173 |
| Body muscle percentage | 1.44 | 2.04 | 1.33 | 1.602 | 0.207 |
| Body water percentage | 2.39 | 3.06 | 2.03 | 1.817 | 0.168 |
| Serum total adiponectin (µg/ml) | 5.34 | 5.03 | 5.54 | 0.046 | 0.955 |
| Fasting serum glucose (mg/dl) | -1.59 | -0.89 | 0.00 | 0.068 | 0.935 |
| Fasting serum insulin (µIU/ml) | -3.19 | -2.36 | -1.35 | 1.013 | 0.367 |
| HOMA-IR | -0.41 | -0.30 | -0.18 | 1.021 | 0.364 |

Abbreviations: F variation between sample means/variation within the samples in the ANOVA test, HOMA-IR Homeostasis model assessment for insulin resistance, N/A Not applicable

being aware of the health hazards of obesity. The duration of the intervention program was the second limitation, after analyzing data; the need for more than 16 weeks of intervention was raised to test the effect of exercise on adiponectin level. This duration was chosen to decrease the number of dropouts and the expectation of dropouts was 10%, but actually, they reached 22%.

5 Conclusion

Lifestyle-related factors, such as overeating and physical inactivity, induce the accumulation of visceral fat which may lead to dysfunction of adipocytes. Hyposecretion of defensive adiponectin might represent obvious mechanisms of lifestyle-related diseases, including diabetes mellitus, hypertension, hyperlipidemia, and atherosclerosis. Reduction of visceral fat is recommended as it can be a major preventive measure for the metabolic syndrome and its consequences, as our study revealed that a moderate weight loss following a balanced low caloric diet was associated with a significant increase in serum total adiponectin level, percent of muscle mass and body water, and a significant reduction in insulin resistance, and waist circumference.

Abbreviations

ANOVA: Analysis of variance; BIA: Bioelectrical impedance analysis; BMI: Body mass index; ELISA: Enzyme-linked immunosorbent assay; HC: Hip circumference; HIPH: High Institute of Public Health; Ht: Height; HOMA: Homeostasis model assessment; HOMA-IR: Homeostasis model assessment-insulin resistance; IL-6: Interleukin-6; IR: Insulin resistance; n: Number; P: P value; REE: Resting energy expenditure; SD: Standard deviation; Sig.: Significance; SPSS: Statistical Package of Social Science; TNF-α: Tumor necrosis factor-α; WC: Waist circumference; WHR: Waist-hip ratio; WHtR: Waist-height ratio; Wt: Weight

Acknowledgements

Not applicable.

Authors' contributions

Study concept and design: WHF, RHE, and FAE. Acquisition, analysis, and interpretation of the data: All authors. Drafting of the manuscript: WHF. Revision of the manuscript: All authors. Statistical analysis: SAM and WHF. Administrative and technical support: WHF, RH, FAS, and MKE. The authors have read and approved the manuscript.

Funding

No funding for this research was received.

Availability of data and materials

The datasets used and analyzed during the current study are available from the corresponding author on reasonable request.

Ethics approval and consent to participate

Approval of the Ethics Committee of HIPH, Alexandria University, Egypt, was obtained on 9 February 2016. Committee's reference number is not applicable. All patients were informed, and a written consent was obtained from all participants after explaining the aim of the study.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Author details

¹Nutrition Department, High Institute of Public Health, Alexandria University, Alexandria, Egypt. ²Chemical Pathology Department, Medical Research Institute, Alexandria University, Alexandria, Egypt. ³Biostatistics Department, High Institute of Public Health, Alexandria University, Alexandria, Egypt.

Received: 23 December 2019 Accepted: 5 October 2020

Published online: 01 December 2020

References

1. World Health Organization. Obesity and overweight. Geneva: WHO; 2015. Fact sheet No.311.

2. Ministry of Health and Population (Egypt), El-Zanaty and Associates ICF International. Egypt Demographic and Health Survey 2014. Cairo, Rockville: Ministry of Health and Population, ICF International; 2015. p. 179.
3. Havel PJ. Control of energy homeostasis and insulin action by adipocyte hormones: leptin, acylation stimulating protein, and adiponectin. *Curr Opin Lipidol*. 2002;13(1):51–9.
4. Frühbeck G, Catalán V, Rodríguez A, Ramírez B, Becerril S, Salvador J, et al. Adiponectin-leptin ratio is a functional biomarker of adipose tissue inflammation. *Nutrients*. 2019;11(2):454.
5. Kadowaki T, Yamauchi T. Adiponectin and adiponectin receptors. *Endocr Rev*. 2005;26(3):439–51.
6. Stefan N, Stumvoll M. Adiponectin—its role in metabolism and beyond. *Horm Metab Res*. 2002;34(09):469–74.
7. Cnop M, Havel PJ, Utzschneider KM, Carr DB, Sinha MK, Boyko EJ, et al. Relationship of adiponectin to body fat distribution, insulin sensitivity and plasma lipoproteins: evidence for independent roles of age and sex. *Diabetologia*. 2003;46(4):459–69.
8. Hu E, Liang P, Spiegelman BM. AdipoQ is a novel adipose-specific gene dysregulated in obesity. *J Biol Chem*. 1996;271(18):10697–703.
9. Kadowaki T, Yamauchi T, Kubota N, Hara K, Ueki K, Tobe K. Adiponectin and adiponectin receptors in insulin resistance, diabetes, and the metabolic syndrome. *J Clin Invest*. 2006;116(7):1784–92.
10. Hotta K, Funahashi T, Arita Y, Takahashi M, Matsuda M, Okamoto Y, et al. Plasma concentrations of a novel, adipose-specific protein, adiponectin, in type 2 diabetic patients. *Arterioscl Thromb Vas*. 2000;20(6):1595–9.
11. Brook RD. Obesity, weight loss, and vascular function. *Endocrine*. 2006;29(1):21–5.
12. Eddy DM, Schlessinger L, Kahn R. Clinical outcomes and cost-effectiveness of strategies for managing people at high risk for diabetes. *Ann Intern Med*. 2005;143(4):251–64.
13. Polak J, Kovacova Z, Holst C, Verdich C, Astrup A, Blaak E, et al. Total adiponectin and adiponectin multimeric complexes in relation to weight loss induced improvements in insulin sensitivity in obese women: the NUGENOB study. *Eur J Endocrinol*. 2008;158:533–41.
14. Polak J, Kovacova Z, Jacek M, Klimcakova E, Kovackikova M, Vitkova M, et al. An increase in plasma adiponectin multimeric complexes follows hypocaloric diet-induced weight loss in obese and overweight premenopausal women. *Clin Sci*. 2007;112(11):557–65.
15. Kopp HP, Krzyzanowska K, Möhlig M, Spranger J, Pfeiffer AF, Scherthaner G. Effects of marked weight loss on plasma levels of adiponectin, markers of chronic subclinical inflammation and insulin resistance in morbidly obese women. *Int J Obes (Lond)*. 2005;29:766–71.
16. Mai S, Walker GE, Brunani A, Guzzaloni G, Grossi G, Oldani A, et al. Inherent insulin sensitivity is a major determinant of multimeric adiponectin responsiveness to short-term weight loss in extreme obesity. *Sci Rep-UK*. 2014;4:5803.
17. Jelliffe DB, Jelliffe EFP, Zerfas A, Neumann CG. Community nutritional assessment, with special reference to less technically developed countries. 2nd ed. London: Oxford University Press; 1989.
18. Kyle UG, Bosaeus I, De Lorenzo AD, Deurenberg P, Elia M, Gómez JM, et al. Bioelectrical impedance analysis—part 1: review of principles and methods. *Clin Nutr*. 2004;23(5):1226–43.
19. David B, Sacks MB. Carbohydrates. In: Burtis CA, Ashwood ER, Bruns DE, editors. *Tietz textbook of clinical chemistry*. Philadelphia: Elsevier & Saunders Company; 2008. p. 389–401.
20. Evena MSSC, Barnarda ND, Mistry J, Sinha MK. Development of a novel ELISA for human insulin using monoclonal antibodies produced in serum-free cell culture medium. *Clin Biochem*. 2007;40:98–103.
21. Wallace TM, Levy JC, Matthews DR. Use and abuse of HOMA modeling. *Diabetes Care*. 2004;27:1487–95.
22. Takemura Y, Ouchi N, Shibata R, Aprahamian T, Kirber MT, Sumner RS, et al. Adiponectin modulates inflammatory reactions via calcitriol receptor-dependent clearance of early apoptotic bodies. *J Clin Invest*. 2007;117:375–86.
23. Lysen LK, Israel DA. Nutrition in weight management. In: Mahan LK, Raymond JL, editors. *Krause's food and the nutrition care process*. St. Louis: Elsevier Inc; 2017. p. 383–406.
24. Mifflin MD, St Joer ST, Hill LA, Scott BJ, Daugherty SA, Koh YO. A new predictive equation for resting energy expenditure in healthy individuals. *Am J Clin Nutr*. 1990;51(2):241–7.
25. Institute of Medicine, Food and Nutrition Board. Dietary reference intakes (DRI) for energy, carbohydrate, fiber, fat, fatty acids, cholesterol, protein, and amino acids (2002/2005). Washington DC: The National Academic Press; 2005. p. 1329.
26. Zar JH. *Biostatistical analysis*. Pearson new international edition, Pearson Higher Ed; 2013.
27. Arawaka N, Daimon M, Oizumi T, Jimbu Y, Kameda W, Yamaguchi H, et al. Correlation between change in body weight rather than current body weight and change in serum adiponectin levels in a Japanese population—the Funagata study. *Metabolism*. 2006;55(3):324–30.
28. Xydakis AM, Case CC, Jones PH, Hoogeveen RC, Liu MY, Smith EO, et al. Adiponectin, inflammation, and the expression of the metabolic syndrome in obese individuals: the impact of rapid weight loss through caloric restriction. *J Clin Endocrinol Metab*. 2004;89(6):2697–703.
29. Ryan AS, Nicklas BJ, Berman DM, Elahi D. Adiponectin levels do not change with moderate dietary induced weight loss and exercise in obese postmenopausal women. *Int J Obes (Lond)*. 2003;27(9):1066–71.
30. Serra A, Granada ML, Romero R, Bayés B, Cantón A, Bonet J, et al. The effect of bariatric surgery on adipocytokines, renal parameters and other cardiovascular risk factors in severe and very severe obesity: 1-year follow-up. *Clin Nutr*. 2006;25(3):400–8.
31. Kotidis EV, Koliakos G, Papavramidis TS, Papavramidis ST. The effect of biliopancreatic diversion with pylorus-preserving sleeve gastrectomy and duodenal switch on fasting serum ghrelin, leptin and adiponectin levels: is there a hormonal contribution to the weight-reducing effect of this procedure? *Obes Surg*. 2006;16(5):554–9.
32. Yamamoto Y, Hirose H, Saito I, Tomita M, Taniyama M, Matsubara K, et al. Correlation of the adipocyte-derived protein adiponectin with insulin resistance index and serum high-density lipoprotein-cholesterol, independent of body mass index, in the Japanese population. *Clin Sci*. 2002;103(2):137–42.
33. Zillikens MC, Uitterlinden AG, van Leeuwen JP, Berends AL, Henneman P, van Dijk KW, et al. The role of body mass index, insulin, and adiponectin in the relation between fat distribution and bone mineral density. *Calcif Tissue Int*. 2010;86(2):116–25.
34. Matsubara M, Maruoka S, Katayose S. Inverse relationship between plasma adiponectin and leptin concentrations in normal-weight and obese women. *Eur J Endocrinol*. 2002;147(2):173–80.
35. Kuo SM, Halpern MM. Lack of association between body mass index and plasma adiponectin levels in healthy adults. *Int J Obes (Lond)*. 2011;35(12):1487–94.
36. Yang WS, Lee WJ, Funahashi T, Tanaka S, Matsuzawa Y, Chao CL, et al. Weight reduction increases plasma levels of an adipose-derived anti-inflammatory protein, adiponectin. *J Clin Endocrinol Metab*. 2001;86(8):3815–9.
37. Lindsay RS, Resnick HE, Zhu J, Tun ML, Howard BV, Zhang Y, et al. Adiponectin and coronary heart disease: the strong heart study. *Arterioscl Thromb Vas*. 2005;25(3):e15–6.
38. Weyer C, Funahashi T, Tanaka S, Hotta K, Matsuzawa Y, Pratley RE, et al. Hypoadiponectinemia in obesity and type 2 diabetes: close association with insulin resistance and hyperinsulinemia. *J Clin Endocrinol Metab*. 2001;86(5):1930–5.
39. Tschritter O, Fritsche A, Thamer C, Haap M, Shirkavand F, Rahe S, et al. Plasma adiponectin concentrations predict insulin sensitivity of both glucose and lipid metabolism. *Diabetes*. 2003;52(2):239–43.
40. Esposito K, Pontillo A, Di Palo C, Giugliano G, Masella M, Marfella R, et al. Effect of weight loss and lifestyle changes on vascular inflammatory markers in obese women: a randomized trial. *JAMA*. 2003;289(14):1799–804.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.